## Package 'DEHOGT'

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Type Package

Title Differentially Expressed Heterogeneous Overdispersion Gene Test for Count Data

Version 0.99.0

Description Implements a generalized linear model approach for detecting differentially expressed genes across treatment groups in count data. The package supports both quasi-Poisson and negative binomial models to handle over-dispersion, ensuring robust identification of differential expression. It allows for the inclusion of treatment effects and gene-wise covariates, as well as normalization factors for accurate scaling across samples. Additionally, it incorporates statistical significance testing with options for p-value adjustment and log2 fold range thresholds, making it suitable for RNA-seq analysis as described in by Xu et al., (2024) <doi:10.1371/journal.pone.0300565>.

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**Encoding** UTF-8

**Depends** R (>= 3.5.0)

Imports doParallel, foreach, MASS,

Suggests knitr, rmarkdown, BiocStyle

**biocViews** GeneExpression, DifferentialExpression, StatisticalMethod, Regression, Normalization

VignetteBuilder knitr

RoxygenNote 7.3.2

URL https://github.com/ahshen26/DEHOGT

BugReports https://github.com/ahshen26/DEHOGT/issues

NeedsCompilation no

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	posed method in the above paper

#### Description

Differentially Expressed Heterogeneous Overdispersion Genes Testing for Count Data This script implements the main function of the proposed method in the above paper

#### Usage

```
dehogt_func(
   data,
   treatment,
   norm_factors = NULL,
   covariates = NULL,
   dist = "qpois",
   padj = TRUE,
   pval_thre = 0.05,
   l2fc = FALSE,
   l2fc_thre = 1,
   num_cores = 1
)
```

#### Arguments

data	A matrix of gene expression data where rows represent genes and columns represent samples.
treatment	A vector specifying the treatment conditions for each sample.
norm_factors	An optional vector of normalization factors for each sample. Default is NULL, which assumes equal normalization factors.
covariates	An optional matrix of gene-wise covariates. Default is NULL.
dist	The distribution family for the GLM. Can be "qpois" for quasi-Poisson or "neg- bin" for negative binomial. Default is "qpois".

#### dehogt\_func

padj	Logical value indicating whether to adjust p-values using the Benjamini-Hochberg (BH) procedure. Default is TRUE.
pval_thre	The threshold for identifying differentially expressed genes based on adjusted p-values. Default is 0.05.
l2fc	Logical value indicating whether to consider log2 fold change for identifying differentially expressed genes. Default is FALSE.
l2fc_thre	The threshold for log2 fold change in identifying differentially expressed genes. Default is 1.
num_cores	The number of CPU cores to use for parallel computing. Default is 1.

#### Value

A list containing:

DE_idx	A logical vector indicating differentially expressed genes.
pvals	A numeric vector of p-values for each gene.
log2fc	A numeric vector of log2 fold changes for each gene.

#### Examples

```
# simulate gene expression data
data <- matrix(rpois(1000, 10), nrow = 100, ncol = 10)
# simulate random treatment assignments
treatment <- sample(0:1, 10, replace = TRUE)
# Run main function with parallel computing using 2 cores
result <- dehogt_func(data, treatment, num_cores = 2)</pre>
```

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 $dehogt_func, 2$